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In vivo oximetry of human bulbar conjunctival and episcleral microvasculature using snapshot multispectral imaging

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Abstract

Multispectral imaging (MSI) is now well established for non-invasive oximetry of retinal blood vessels, contributing to the understanding of a variety of conditions affecting the retinal circulation, including glaucoma, diabetes, vessel occlusion, and auto-regulation. We report the application of a unique snapshot MSI technique to enable the first oximetric imaging of the blood vessels of the anterior segment, i.e. the episcleral and bulbar conjunctival microvasculature. As well as providing a new capability of oximetry of the scleral vasculature, this technique represents ocular oximetry that is complimentary or alternative to retinal oximetry. We report the oxygen dynamics of these microvascular beds and assess how acute mild hypoxia effects the blood oxygen saturation (SO_2) of bulbar conjunctival and episcleral microvasculature.

A retinal-fundus camera fitted with a custom Image-Replicating Imaging Spectrometer enabled oximetric imaging of bulbar conjunctival and episcleral microvasculature in ten healthy human subjects at normoxia (21% Fraction of Inspired Oxygen [FiO_2]) and acute mild-hypoxia conditions (15% FiO_2). Eyelid closure was used to block oxygen diffusion between ambient air and the sclera surface. Four of the ten subjects – those that presented suitable vasculature for direct comparison between bulbar conjunctival and episcleral vessels - were imaged for 30 seconds following eyelid opening. Vessel diameter and Optical Density Ratio (ODR: a direct proxy for oxygen saturation) of vessels was computed automatically. Oximetry capability was validated using a simple phantom for the scleral vasculature,

Average episcleral diameter increased from $78.9 \pm 8.7\mu m$ (mean \pm standard deviation) at normoxia to $97.6 \pm 14.3\mu m$ at hypoxia ($p = 0.02$). Diameters of bulbar conjunctival vessels showed no significant change from $80.1 \pm 7.6\mu m$ at normoxia to

80.6 ± 7.0µm at hypoxia (p= 0.89). Acute mild hypoxia resulted in a decrease in SO₂ (i.e. an increase in ODR) from normoxia levels in both bulbar conjunctival (p <0.001) and episcleral vessels (p= 0.03).

Hypoxic bulbar conjunctival vasculature rapidly re-oxygenated in an exponential manner, reaching normoxia baseline levels, with an average ½ time to full reoxygenation of 3.4 ±1.4 seconds. This reoxygenation occurs because the bulbar conjunctival vessels are in direct contact with ambient air. This is the first study to characterise and also to image the oxygen dynamics of bulbar conjunctival and episcleral microvasculature, and to directly observe the rapid reoxygenation of hypoxic bulbar conjunctival vessels when exposed to air.

Oxygen diffusion into the bulbar conjunctiva must be taken into account to provide meaningful oximetry because bulbar conjunctival vessels will be highly oxygenated (close to 100% SO₂) when exposed to ambient air.

Oximetry of bulbar conjunctival vessels could potentially provide insight into conditions where oxygen dynamics of the microvasculature are not fully understood, such as diabetes, sickle-cell diseases, and dry-eye syndrome. Further, in vivo oximetry of individual capillaries and groups of flowing red blood cells could be achieved with a high magnification slit lamp adapted for MSI.

Keywords: multispectral imaging, oximetry, hypoxia, bulbar conjunctiva, episclera, oxygen saturation, microvasculature, oxygen diffusion,

1. Introduction

Multispectral imaging (MSI) is well established for non-contact oximetry of blood vessels (D J Mordant et al., 2011a; David J Mordant et al., 2011b) which has enhanced the understanding of a variety of retinal conditions, such as diabetes (Hammer et al., 2009; [Hardarson and Stefánsson, 2012](#); Isenberg et al., 1986), glaucoma ([Boeckaert et al., 2012](#); Mordant et al., 2014; [Olafsdottir et al., 2011](#)), and vessel occlusion (Eliasdottir et al., 2014), as well as auto-regulation response to flicker stimulation (Hammer et al., 2011) and acute mild hypoxia (Choudhary et al., 2013). However, oximetry of capillaries in the retina is beyond the technical capabilities of MSI-enabled retinal fundus cameras. The anterior segment provides two alternative ocular microvascular beds that are easily accessible for multispectral imaging and which could be used to probe ocular blood oxygen saturation and potentially provide new physiologically-relevant information; the bulbar conjunctival and episcleral microvascular beds. This is the first study to use MSI to non-invasively measure the oxygen saturation of bulbar conjunctival and episcleral microvasculature with high spatial and temporal resolution, revealing rapid oxygen diffusion from ambient air into bulbar conjunctival vessels.

The episcleral microvasculature is located within the scleral tissue, with few episcleral vessels visible near the scleral surface. In contrast, the bulbar conjunctival microvasculature is semi-mobile above the sclera, and presents many arterioles, venules, and capillaries for imaging ([Meighan, 1956](#)). Groups of individual red blood cells can be observed to flow in bulbar conjunctival capillaries if imaged with high magnification (Jiang et al., 2014). The bulbar conjunctiva may be unique in that it is the only microvascular bed in the human body which is directly exposed to ambient air. Figure 1a shows generalised vessel positions with respect to the sclera. Figure 1b shows a representative image of bulbar conjunctival and episcleral vasculature in

a single subject. However, despite potential for new oximetry information and ease of imaging, no MSI oximetry studies of either the bulbar conjunctival or episcleral microvasculature have been published to date.

MSI oximetry is based on the SO_2 -dependent optical absorption spectra of haemoglobin. Changes in SO_2 can be calculated by imaging blood vessels at two wavelengths: one wavelength where optical absorption is sensitive to variations in SO_2 , and at another wavelength which is insensitive to SO_2 variations (i.e. isobestic). From images of vessels, the optical density (OD) of vessels at each wavelength can be calculated, allowing the calculation of optical density ratio (ODR); ODR is directly proportional to SO_2 . In vessels where SO_2 is known, ODR can then be empirically calibrated to SO_2 by assuming local arterial SO_2 is equal to the SO_2 of systemic arterial SO_2 as measured by pulse oximetry (Beach et al., 1999), or by using reference values from previous studies. (Hardarson et al., 2006).

To the best of our knowledge there are no reported MSI oximetry studies of the bulbar conjunctival or episcleral microvasculature. Instead, insights into the oxygen dynamics of microvasculature have generally been indirectly inferred from vessel-diameter or blood-flow measurements (Jiang et al., 2013; Shahidi et al., 2010; Wanek et al., 2013), however these parameters may be affected by factors other than changes in SO_2 , such as conjunctival or episcleral inflammation. Direct measurement of the partial pressure of oxygen (pO_2) of the palpebral conjunctival microvasculature has been achieved with Clark-type electrodes (Chapman et al., 1986; Iguchi et al., 2005; Isenberg et al., 2002; Kwan and Fatt, 1971; Mader et al., 1987), however these electrodes have insufficient spatial discrimination for localisation of oximetry to blood vessels and crucially, block oxygen diffusion between ambient air and blood vessels under study.

130 In this study, we report the use of a retinal fundus camera modified for Snapshot
131 Multispectral Imaging (SMSI) to non-invasively quantify the oxygen dynamics of both
132 bulbar conjunctival and episcleral microvasculature in ten healthy human subjects.
133 The high temporal resolution of the SMSI system (10ms exposure, 1Hz image
134 acquisition rate) enables observation of fast biological processes (Fernandez
135 Ramos et al., 2014). We observe rapid oxygen diffusion from ambient air into bulbar
136 conjunctival vessels due to the unique location of the bulbar conjunctiva (i.e. directly
137 in contact with ambient air); such observations are not possible with time-sequential
138 MSI or Clarke-type electrodes because these techniques lack sufficient temporal and
139 spatial resolution respectively.

140

141 **2. Material and methods**

142 **2.1. Subject recruitment**

143 This study was approved by the Ethics Committee of the University of Glasgow,
144 College of Medical, Veterinary and Life Sciences. All volunteers provided written
145 informed consent before participation and all procedures were performed in
146 accordance with the tenets of the Declaration of Helsinki. Ten healthy volunteers
147 (age 25 ± 2 years, six males and four female) were recruited. Subjects reported no
148 history of ocular, respiratory, or vascular disease. Volunteers that regularly wore
149 contact lenses or who were suffering from allergic conjunctivitis were excluded
150 because this may induce fluctuating bulbar conjunctival vasodilatation (Gartner,
151 1944; Cheung et al., 2012; Jiang et al., 2014) .

152

153 **2.2. Imaging system**

154 The imaging system consisted of a commercial retinal fundus camera (Topcon
155 TR50-DX; Topcon, Itabashi, Tokyo, Japan), fitted with an Image Replicating Imaging

Spectrometer (IRIS) and a cooled sCMOS camera (Zyla 5.5; Andor, Belfast, United Kingdom). IRIS is discussed in detail elsewhere (Harvey et al., 2005; Alabboud et al., 2007; [Gorman et al., 2010](#); [Fernandez Ramos et al., 2014](#)); but in brief, IRIS simultaneously spectrally de-multiplexes a white-light image into eight distinct narrowband spectral images onto a single detector without rejection of light. Orthogonal-polarization imaging was used to minimise specular reflections from the sclera and blood vessels ([van Zijderveld et al., 2014](#)). Fundus-camera flash and image acquisition were synchronized using a custom graphical user interface written in LabVIEW, and images were saved in uncompressed Tiff format. Image acquisition was limited to 1Hz by the fundus camera flash refresh rate with an exposure time of 10ms. This imaging set-up and a representative multispectral IRIS image of the sclera are shown in Figure 2.

The curved scleral surface presents a challenge for imaging because it causes the position of blood vessels to vary with respect to the imaging plane of the fundus camera, potentially up to ~12mm from the anterior segment to the extreme lateral side of the sclera. To insure sharp focus over an extended scleral region, the 'small aperture' setting of the fundus camera was selected. This resulted in an estimated depth-of-field (DOF) of ~10mm; DOF was estimated by imaging a USAF test chart (USAF 1951 Chart; Applied Image Group-Imaging, Rochester, New York, USA) as it was moved through prime-focus on a linear-translation stage. A 35-degree field-of-view was selected to provide a field of view at the object plane of approximately 85 x 45mm. This combination of settings enabled the imaging of bulbar conjunctival and episcleral vessels over an extended scleral region with an optimal, sharp focus.

2.3. Scleral phantom

For assessment of the validity of our oximetry technique, a simple sclera-mimicking phantom was manufactured (see Figure 3). Similar phantoms have previously been used to validate retinal oximetry (David J Mordant et al., 2011). The phantom consisted of a transparent Fluorinated Ethylene Propylene (FEP) capillary of 100µm inner diameter (Zuess inc., Belfast, Northern Ireland), placed in contact with optical-grade Spectralon (Spectralon® Diffusion Material; Labsphere inc, North Sutton, New Hampshire, USA); Spectralon has similar spectral reflectance characteristics to the sclera (Bashkatov et al., 2010; Labsphere Inc.). To simulate in vivo blood circulation, ex vivo whole horse blood (40% hematocrit) (E&O labs, Bonnybridge, Scotland, United Kingdom) was flowed through the FEP capillary under feed from a syringe pump (KDS260, Linton Instrumentation, UK). SO₂ of the blood was reduced by adding measured quantities of Sodium Dithionite (EMD Millipore, Fisher Scientific, Loughborough, UK) to 5ml samples of blood according to the procedure described in Briely-Sabo and Bjornerud (Briley-Saebo and Bjornerud, 2000). SO₂ blood samples was measured prior to imaging using an optical blood gas analyser (GEM OPL, Instrumentation Laboratory, Bedford, Massachusetts, USA). A total of eight SO₂ samples ranging between 5% and 100% SO₂ were imaged in the FEP capillary.

2.4.1. Experimental procedure for in-vivo imaging

~~Subjects positioned their head in the standard fundus-camera chin-rest; head-straps were used to restrain the subject and minimise any motion. The fundus camera objective lens was positioned approximately five centimetres from the subject's sclera. In this configuration, the fundus camera illumination formed a circle approximately four centimetres in diameter. Subject gaze was controlled by the subject fixating on the fundus camera external fixation target (a movable red LED). For each subject, the scale of images was calibrated by imaging a millimeter scale located in the nominal plane of the sclera at prime focus. This yielded an average~~

~~image scale of 13.5 microns per pixel, enabling conversion of vessel diameter in pixels to diameter in microns.~~

Subjects positioned their head in the standard fundus-camera chin-rest; head-straps were used to restrain the subject and minimise any motion. The fundus camera objective lens was positioned approximately five centimetres from the subject's sclera. In this configuration, the fundus camera illumination formed a circle approximately four centimetres in diameter. Subject gaze was controlled by the subject fixating on the fundus camera external fixation target (a movable red LED). For each subject, the scale of images was determined by imaging a millimeter scale placed in front of the sclera at prime focus. All subsequent images were acquired at this focal position. This enabled a calibration of the size of each pixel on the detector to the real size of an image; on average, one pixel corresponded to ~13.5 microns. From this, the measured vessel diameter in pixels was calibrated to diameter in microns."

Scleral regions of each subject were selected for imaging so as to maximise the number of bulbar conjunctival vessels meeting the inclusion criteria (see Section 2.6.1). Bulbar conjunctival and episcleral vasculature was distinguished by moving the gaze of a subject; this moved the position of the bulbar conjunctiva above the sclera, altering the relative position of bulbar conjunctival and episcleral vessels. However it was not possible to classify individual vessels as arterioles and venules because of the diverse morphology of bulbar conjunctival vasculature and the limited number of episcleral vessels available for imaging (see Section 4.4). Scleral regions were chosen for imaging so as to maximise the number of bulbar conjunctival vessels meeting inclusion criteria whilst including some episcleral vessels for analysis (see Section 2.6.1). Once selected, the same blood vessels in a single

scleral region of a single eye for each subject were consistently imaged and analysed throughout the experiment.

Throughout the imaging protocol, the scleral region exposed to air was kept constant by the subject constantly gazing at the stationary fixation target and peripheral arterial SO_2 was recorded throughout the experiment using a fingertip pulse oximeter (AUTOCORR; Smiths Medical ASD Inc., Rockland, MA, USA) interfaced to a computer using a custom LabVIEW interface.

2.4.2. Repeatability

To assess repeatability of ODR measurement, eight consecutive images of the same scleral region were acquired in a period of approximately ten seconds for each subject. Gaze fixation was maintained for 2.5 minutes with their eyelid open prior to imaging to expose the target vasculature to ambient air.

2.4.3. Effect of eyelid closure

Eyelid closure was used to control oxygen diffusion; eyelid closure places a tissue barrier between the scleral surface and the ambient air, drastically decreasing the rate of any oxygen diffusion from ambient air to this scleral surface. To assess if eye closure affects the ODR of vessels, subjects were imaged before and after a period of eyelid closure. As before, subjects continually gazed at the fixation target for 2.5 minutes to expose the target vasculature to ambient air prior to imaging; subjects then closed their eyelids for a further 2.5 minutes. After 2.5 minutes of eyelid closure subjects opened their eyelid and synchronised imaging occurred

2.5.4. Acute mild hypoxia

To assess the effects of acute mild hypoxia on ODR, subjects were imaged at normoxia and acute mild-hypoxia. For normoxia measurement, subjects inhaled room air (21% FiO₂) for 2.5 minutes whilst fixating on the red LED fixation target, after which they were imaged. To induce acute mild hypoxia, subjects closed their eyelids and breathed a hypoxic air mixture (15% 2.5 minutes of inhalation of hypoxic air mixture (15% FiO₂) supplied via a hypoxic-air generator (Everest Summit II Hypoxic Generator; Hypoxico, Inc., New York, NY, USA) (Spurling et al., 2011). The hypoxic-air generator was calibrated before use and the air supply was monitored with an in-line oxygen analyzer (AD300 oxygen analyser; Teledyne Analytical Instruments, City of Industry, California, USA). Hypoxic air generators have been previously used for a study into retinal response to acute mild hypoxia (Choudhary et al., 2013).

After 2.5 minutes of hypoxic-air inhalation, subjects opened their eyelids and synchronised imaging occurred. Synchronisation of imaging with events, such as eyelid opening, was accomplished with a five-second oral countdown and with an accuracy of ± 1 seconds. Subjects were then returned to normoxia by breathing room air. This process was repeated in the following sequence: normoxia 1, hypoxia 1, normoxia 2, hypoxia 2, normoxia 3; this sequence provides a robust time-sequential modulation in SO₂ and associated ODR change that is highly distinct from normal physiological variations.

2.5.5. Exposure of hypoxic vasculature to ambient air

A sub-group of four subjects (3 male, 1 female) were selected for further study. These subjects presented bulbar conjunctival and episcleral vessels suitable for analysis within single scleral region, allowing concurrent imaging - and thus comparison of oxygen dynamics - between bulbar conjunctival and episcleral

vessels. Hypoxia was induced as described in section 2.5.3. However, when subjects opened their eyelids, a synchronised 1Hz frame-rate imaging sequence was subsequently recorded for the 30 seconds, enabling observation of any rapid diffusion processes. This was repeated twice per subject.

2.6. Image analysis

2.6.1 Vessel section inclusion criteria

The following inclusion criteria were applied to ensure that only appropriate vessel sections were selected for analysis: (1) vessel sections had to be greater than 5 pixels ($\sim 67\mu\text{m}$) in diameter to ensure that the contrast is not significantly affected by the modulation-transfer function of the imaging system. (2) vessel section had to have no other vessel sections within 12 pixels of either side of the vessel to be analysed; the presence of small vessels was accepted due to the high number of small bulbar conjunctival vessels; (3) vessel sections had to be at least 30 pixels long ($\sim 405\mu\text{m}$); (4) vessels close to vessel intersections, regions of scleral glare, specular reflections, or images with poor focus were excluded; (5) episcleral vessels had to be of high apparent contrast with respect to the scleral tissue and not show a significant decrease in contrast along the analysed vessel section length (i.e. not appear to go deeper in the sclera tissue); (6) vessel sections had to meet all these inclusion criteria for all images in each section of the study.

2.6.2. Vessel tracking

Image processing was implemented post hoc using custom algorithms implemented in MATLAB. Raw IRIS images were cropped and co-registered to create a multispectral data cube. Vessels were tracked semi-automatically using manually identified control points. Repeated semi-automatic tracking demonstrated negligible variation in ODR (a standard deviation of $<0.5\%$ in 10 repeated measurements).

Fully automatic tracking was not implemented because inter-image registration of bulbar conjunctival vessels is affected by the relative motion of bulbar conjunctival and episcleral vasculature (Crihalmeanu and Ross, 2012).

2.6.3. Oximetric analysis and vessel diameter measurement

Our oximetric analysis is based on two-wavelength oximetry developed by Beach et al (Beach et al., 1999). For two-wavelength oximetry, the optical-density (OD) of blood vessels at two spectral wavebands is calculated: one waveband where optical absorption is insensitive to changes in SO₂ (isobestic) and one waveband where optical absorption is sensitive to changes in SO₂ (contrast). The 570nm IRIS waveband was utilised as the isobestic reference and the 560nm waveband was used as the oxygen sensitive waveband (Prahl, 1999). Each waveband has a full spectral-width of approximately 7nm (Fernandez Ramos et al., 2014). Simple modelling based upon the Beer-Lambert law of optical absorption shows that the OD of blood vessels of ~60-100µm at 560nm and 570nm wavebands is expected to be between 0.15 and 1; near-optimal for oximetry (van Assendelft, 1970).

A vessel-fitting algorithm was used to estimate vessel diameter (in pixels) and optical transmission of vessels (see Figure 4). Vessel diameter at 570nm was estimated according to the method described by Fischer et al., (Fischer et al., 2010), where the vessel boundaries are defined as the points in the vessel profile with the maximum rate of change in grayscale intensity. This provided reputable fitting for both bulbar conjunctival and episcleral vessels. Using this fitting algorithm, greyscale intensity in the centre of each vessel (I_v) was calculated and the background greyscale intensity at the centre of the vessel (I_o) was estimated by a linear fit to the background. OD was then calculated for each wavelength by:

$$OD_{\lambda} = -\log_{10} \left(\frac{I_v}{I_o} \right). \quad (1)$$

ODR, defined as $ODR = OD_{560}/OD_{570}$, was then calculated for each vessel; ODR is a direct proxy for SO_2 ; if SO_2 increases, ODR decreases. ODR is approximately independent of vessel diameter and concentration of hemoglobin.

If two or more reference SO_2 values are known, then ODR can be empirically calibrated to SO_2 (Beach et al., 1999). However, no calibration is possible for this study because no empirical measurements of SO_2 in either bulbar conjunctival or episcleral vasculature have been reported in the literature, so we report results simply in terms of ODR.

3. RESULTS

3.1 Sclera phantom

A total of eight ex vivo blood samples of various oxygenations were imaged and analysed in the scleral phantom. Some variation in ODR was seen as blood flowed along the capillary. Overall, ODR was found to decrease with increasing SO_2 and the data was well fitted by a linear trend ($R^2 = 0.89$) (see Figure 5), validating the use of our MSI technique for oximetry of vessels in a scleral-like configuration. Repeatability of scleral phantom ODR measurements was $<0.5\%$ (standard deviation of 10 consecutive images).

3.2 Repeatability of in vivo ODR

The repeatability of in vivo ODR measurements is summarised in Table 1. The greater repeatability of ODR measurement of bulbar conjunctival vessels (0.96%) compared to episcleral vessels (1.55%) when calculated as an average across vessel type is probably due to the larger number of bulbar conjunctival vessel sections analyzed (57 in total) compared to episcleral vessel sections (22 in total); the larger number of vessels analysed reduces the sensitivity to fluctuations in ODR.

369

370 **3.4. Eyelid closure during normoxia**

371 Eyelid closure during normoxia resulted in no statistically significant change in ODR
372 of either bulbar conjunctival or episcleral vessels. When the eyelid was open with
373 constant gaze for 2.5 minutes, the average ODR was 0.90 ± 0.08 (mean \pm standard
374 deviation) for bulbar conjunctival vessels and 0.94 ± 0.09 for episcleral vessels. After
375 eyelid closure, average ODR was 0.90 ± 0.08 for bulbar conjunctival vessels and
376 0.93 ± 0.08 for episcleral vessels ($p = 0.99, 0.72$ respectively; paired t-test).

377

378 **3.5. Acute mild hypoxia**

379 Table 2 and Figure 6 summarise measurements of ODR, vessel diameter, and
380 fingertip pulse oximetry at normoxia and hypoxia. Figure 6a shows ODR and pulse
381 oximeter data throughout the whole normoxia/hypoxia sequence. Bulbar
382 conjunctival ODR increased with hypoxia (indicating a reduction in SO_2) from $0.846 \pm$
383 0.014 (mean \pm standard error) at normoxia to 0.916 ± 0.011 at hypoxia ($p < 0.001$,
384 paired t-test) (Figure 6b). Episcleral ODR increased on average, from 0.881 ± 0.019
385 (mean \pm standard error) at normoxia to 0.938 ± 0.018 at hypoxia ($p = 0.03$, paired t-
386 test) (Figure 6c). Figure 7 shows an overlaid ODR map of vessels at normoxia and
387 hypoxia.

388

389 Bulbar conjunctival vessel diameter did not change significantly between normoxia
390 and hypoxia ($p = 0.89$, paired t-test), however increases in vessel diameters were
391 apparent in some subjects, whereas decreases in diameters were seen in others
392 (Figure 6d). Diameters of episcleral vessels were observed to increase from $78.9 \pm$
393 $8.65\mu m$ (mean \pm standard deviation) at normoxia to $97.6 \pm 14.3\mu m$ at hypoxia
394 (Figure 6e) ($p = 0.02$, paired t-test).

395

3.6. Exposure of hypoxic vasculature to ambient air

For all eight datasets (four subjects, each imaged twice) ODR of hypoxic bulbar conjunctival vessels rapidly decreased upon eyelid opening (indicating an increase in SO_2), tending asymptotically to an ODR corresponding to ODR measured at normoxia. The variation in ODR was well-fitted by an exponential-decay function representing re-oxygenation of the conjunctival vessels plus a linear component, reflecting the incoming hypoxic blood supply:

$$OD = a * e^{-bt} + ct + d \quad (2)$$

Where t is time and a , b , c , d , are empirically calculated constants. The half-time to full reoxygenation ($T_{1/2}$) can then be calculated by:

$$T_{1/2} = -\frac{\ln(2)}{b}. \quad (3)$$

$T_{1/2}$ varied on both an intra and inter-subject basis (see Table 3) but averaged over all measurements $T_{1/2}$ was 3.4 ± 1.4 seconds (mean \pm standard deviation). Figure 8 shows this reoxygenation process in two representative subjects.

Episcleral vessel ODR remained higher (i.e. lower SO_2) after eyelid opening than at normoxia levels and was well fitted by a linear trend. Pulse oximeter SO_2 followed a similar trend to episcleral ODR.

4. Discussion

4.1. Validation of oximetry technique using scleral phantom

Results from the scleral phantom measurement validated the ability of the spectral imaging technique to characterise ODR for oximetry for blood vessels. Some variation in ODR was observed when blood flowed through the capillaries; this variation is likely to be due to non-homogenous SO_2 due to non-uniform deoxygenation by discrete crystals of Sodium Dithionite added to blood (Briley-Saebo and Bjornerud, 2000). Further variation in ODR may be caused by the

aggregation of blood cells, which alters the optical path of light through blood.

Nevertheless, the results shown in Figure 5 clearly support that ODR decreases with SO_2 .

4.2. Effects of acute mild hypoxia

In episcleral vessels, vessel diameter increased and SO_2 decreased at acute mild hypoxia conditions. This is similar to auto-regulation of retinal vessels during acute mild hypoxia (Choudhary et al., 2013). In bulbar conjunctival vessels, SO_2 also decreased with hypoxia, but average vessel diameter did not change significantly. This confirms that the increase in ODR observed is due a decrease in SO_2 and not due a secondary effect due to change of vessel diameter.

4.3. Consequences of oxygen diffusion

Our study is the first to directly show that oxygen diffusion from air results in rapid reoxygenation and saturation of hypoxic bulbar conjunctival vessels. This measurement would not be possible with Clark electrodes, which are limited to a single point measurement and crucially, block oxygen diffusion between ambient air and the tissue in measurement. Hill and Fatt (1963) did however use a Clarke electrode to demonstrate that the bulbar conjunctiva would uptake oxygen from a limited pO_2 reservoir via diffusion, concluding that oxygen diffusion from ambient air to the exposed bulbar conjunctival vessels occurs constantly (Hill and Fatt, 1963). This study is the first to directly observe how this oxygen diffusion alters bulbar conjunctival SO_2 .

It is expected that when in equilibrium with ambient air ($pO_2 \sim 160\text{mmHg}$), bulbar conjunctival vessels will be close to 100% SO_2 because normal arterial blood ($\sim 95\text{-}97\% SO_2$) corresponds to a typical pO_2 of 80-100mmHg; much less than 160mmHg

(Verma and Roach, 2010; Williams, 1998). The average ODR of exposed bulbar conjunctival vessels was consistently ~0.95 (see Figure 6a), indicating a constant equilibrium as expected.

In retinal oximetry, ODR is often empirically calibrated to SO_2 by assuming retinal arterial SO_2 to be equal to the systemic arterial SO_2 as measured by a pulse oximeter. Our results show that the oxygen dynamics of episcleral vessels are similar to pulse oximetry, so this calibration approach would be valid for episcleral vessels if arteries and veins could be accurately identified. However, this calibration approach would not be valid for bulbar conjunctival vessels because our results show that the oxygen dynamics of bulbar conjunctival vessels do not reflect the oxygen dynamics of systemic arterial SO_2 .

4.4. Challenges of bulbar conjunctival and episcleral oximetry

In the retina, oximetry results are often reported independently for arterioles and venules. However, in this study we report results for generalised vasculature and not separately as arterioles and venules for several reasons. (1) Bulbar conjunctival arterioles and venules could not be reliably distinguished from morphology alone due to the significant variation in bulbar conjunctival vessel morphology ([Meighan, 1956](#)). (2) Bulbar conjunctival vessels will be highly oxygenated when exposed to ambient air due to oxygen diffusion from air, and thus could not be distinguished on the basis of ODR. (3) The relatively low number of episcleral vessels that met inclusion criteria did not allow sufficient comparison to identify arteries and veins by either vessel morphology or ODR. Reliable discrimination between episcleral arteries and veins could however be achieved with fluorescence angiography ([Ormerod et al., 1995](#)).

Rattlesnaking - a false apparent change in ODR along the length of a vessel section - is a common artefact in two-wavelength oximetry. Rattlesnaking was observed in both bulbar conjunctival and episcleral vessels and can be seen in Figure 7. Rattlesnaking may be caused by a number of factors such as nearby vessels, variations in scattering properties of background tissue, and groups of erythrocytes flowing in vessels. In small vessels, rattlesnaking may be enhanced in magnitude by the small numbers of red blood cells flowing through narrow vessels.

In this study, only the short-term repeatability of oximetry measurements was assessed. In future, quantification of repeatability of measurements over the course of an entire day is desirable to assess longer term variations including fluctuating diurnal variation in vessel diameter and temperature of bulbar conjunctival vessels ([Duench et al., 2007](#)).

4.5. Influence of light scattering by scleral tissue

Optical scattering of light by tissue may influence ODR and vessel diameter measurement. We assume negligible scattering for the bulbar conjunctival vasculature, which lies within a thin ($\sim 33\mu\text{m}$), transparent bulbar conjunctiva (Efron et al., 2009). However, episcleral vessels are embedded in scleral tissue; this will affect our measurement in two ways. Firstly, the sharpness of vessel boundaries may be decreased, which may produce a small increase systematic and random errors in the measurement of vessel diameter for episcleral vessels. The relative change in vessel diameter measured will however be relatively unaffected by scattering. Secondly, scattering from overlying tissue will act to reduce contrast of vessels, generally acting to reduce the changes in ODR observed. Secondly, scattering from overlying tissue will act to reduce contrast of vessels, generally acting to reduce the changes in ODR observed. Scattering will also be increased if

vessels dilate; this will reduce the apparent change in ODR of episcleral vessels which were observed to dilate significantly (see Figure 6e). In the scleral phantom, the FEP plastic of the capillary will contribute to scattering. The challenge of light scattering by tissue and within blood and the absence of reliable SO_2 values for calibration, makes absolute oximetry in bulbar conjunctival and episcleral vessels challenging, however, as we describe here, changes in SO_2 can be robustly characterised with ODR and can provide useful biological insight.

4.6. Future work

There are good prospects of achieving an absolute oximetry, with minimal requirement for calibration by incorporating the modified Beer-Lambert law (Delpy et al., 1988; [Pittman and Duling, 1975](#)) into multi-waveband optical transmission models. Absolute oximetry would be of particular use because there have been no reference values for SO_2 of the bulbar conjunctival or episcleral microvasculature reported in the literature, so two wavelength oximetry cannot be accurately calibrated.

With appropriate flash illumination, imaging at 100Hz or greater could be achieved and oximetry in smaller bulbar conjunctival vessels and capillaries could be enabled by adapting a slit lamp for high-magnification multispectral imaging. This could enable the potential for non-contact oximetry of groups of red blood cells in humans in vivo. Individual red blood cell oximetry has previously been achieved ex vivo using SMSI ([Fernandez Ramos et al., 2014](#)) and invasively in vivo in anaesthetised mice by photoacoustic microscopy (Wang et al., 2013). SMSI offers faster image acquisition and a simpler image system compared to PAM.

4.7. Vascular conditions that may affect anterior segment vessel SO_2

Understanding SO_2 of bulbar conjunctival and episcleral vessels may provide insight into a range of conditions. For example, diabetic retinopathy is known to result in increased retinal vessel SO_2 (Hammer et al., 2009; Hardarson and Stefánsson, 2012), however, previous studies have shown that oxygen tension in diabetic subjects is lower than in healthy controls (Isenberg et al., 1986). Further, diabetes is associated with increased bulbar conjunctival vessel diameter (Cheung, Anthony T. W. Ramanujam et al., 2001), capillary loss (Owen et al., 2008), and decreased vessel reactivity (Fenton et al., 1979). Snapshot multispectral-imaging oximetry could also provide direct in vivo measurement of resultant hypoxia in bulbar conjunctival vasculature from contact lens wear (Heitmar et al., 2012; Sweeney, 2013). Furthermore oximetry of the bulbar conjunctival vessels may be of interest in studying the recovery of ocular burns using oxygen therapy (Sharifipour et al., 2011), recovery of circulation after surgical or traumatic wound healing, and possibly in the study of ischemic conditions such as dry-eye syndrome (Menezo and Lightman, 2004). High intra-ocular pressure (IOP) is associated with narrowed episcleral veins and increased diameter of episcleral arteries (Nanba and Schwartz, 1986), but it is not known if this may alter episcleral SO_2 .

5. Conclusions

This is the first study to quantify changes localised in SO_2 of bulbar conjunctival and episcleral microvasculature. Oximetry was achieved using SMSI and was validated using a sclera-mimicking phantom.

In vivo, acute mild hypoxia resulted in a repeatable reduction in SO_2 of both bulbar conjunctival and episcleral microvasculature. Episcleral vessels were observed to dilate due to acute mild hypoxia, whereas bulbar conjunctival vessels did not show

statically significant dilation under hypoxia. Hypoxic bulbar conjunctival vessels were observed to rapidly reoxygenate due to oxygen diffusion when exposed to ambient air. Episcleral vessels were not observed to reoxygenate due to overlying episcleral tissue. This oxygen diffusion means that after exposure to air, the pO₂ of bulbar conjunctival vessels will be in equilibrium with ambient air, resulting in a SO₂ close to 100%. SMSI is currently the only oximetry technique with sufficient spatiotemporal resolution to measure this rapid oxygen diffusion in individual vessels. However we have shown that the role of oxygen diffusion in the bulbar conjunctiva must be considered for any future oximetry studies to provide meaningful results.

SMSI oximetry of the bulbar conjunctival and episcleral microvasculature may be of interest in investigating oxygen dynamics in a variety of microvasculature conditions where hypoxia may play a role, such as diabetes, (Isenberg et al., 1986; Hammer et al., 2009; [Hardarson and Stefánsson, 2012](#)), sickle-cell disease (Isenberg et al., 1987), dry-eye syndrome ([Menezo and Lightman, 2004](#)), contact lens wear (Heitmar et al., 2012; [Sweeney, 2013](#)), high intra-ocular pressure (Nanba and Schwartz, 1986), traumatic or surgical wound healing, and ocular-burn recovery (Sharifipour et al., 2011). Further, high-magnification MSI of the bulbar conjunctiva could enable non-invasive in vivo oximetry of individual red blood cells.

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746

Figure 1 (a) Simplified diagram showing position of bulbar conjunctival and episcleral vasculature with respect to the sclera and ambient air. **(b)** Representative image of vasculature observed when imaging the sclera. Bulbar conjunctival vessels are marked with white arrows and episcleral vessels are marked with yellow dashed diamond arrows.

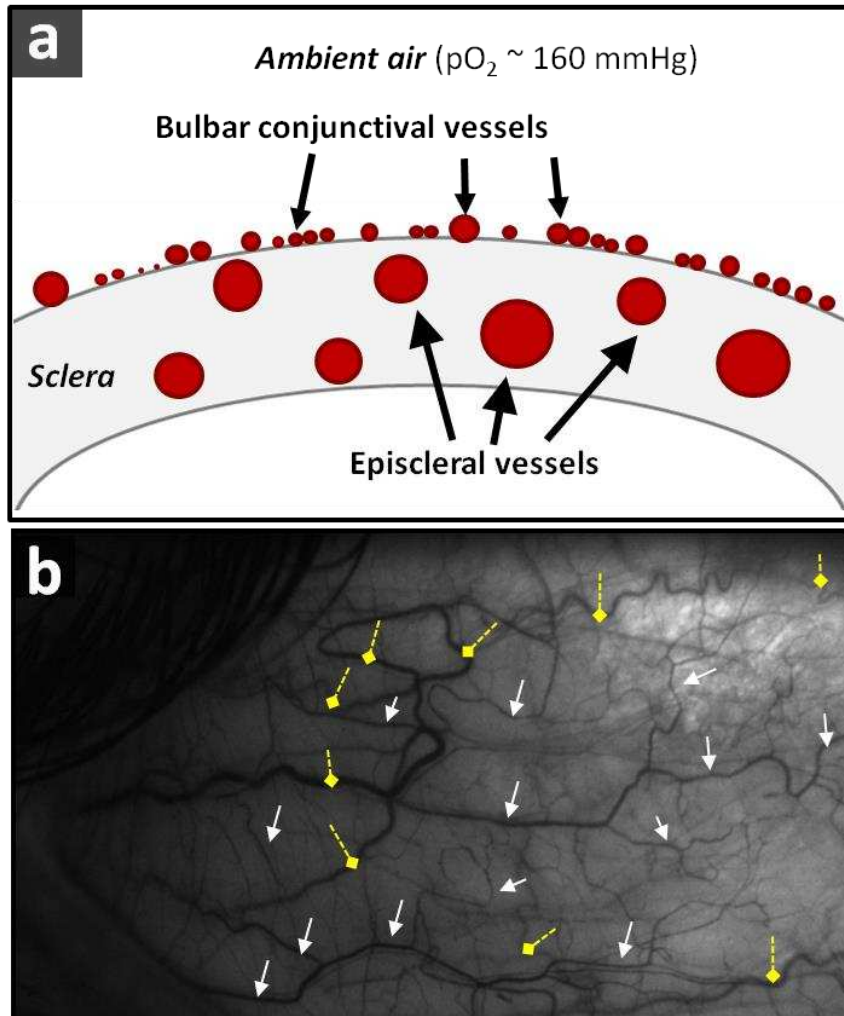


Figure 2 (a) The imaging system: a commercial fundus camera with the Image Replicating Imaging Spectrometer (IRIS) fitted to the upper imaging port. **(b)** A representative 8-band IRIS image of bulbar conjunctival and episcleral vasculature.

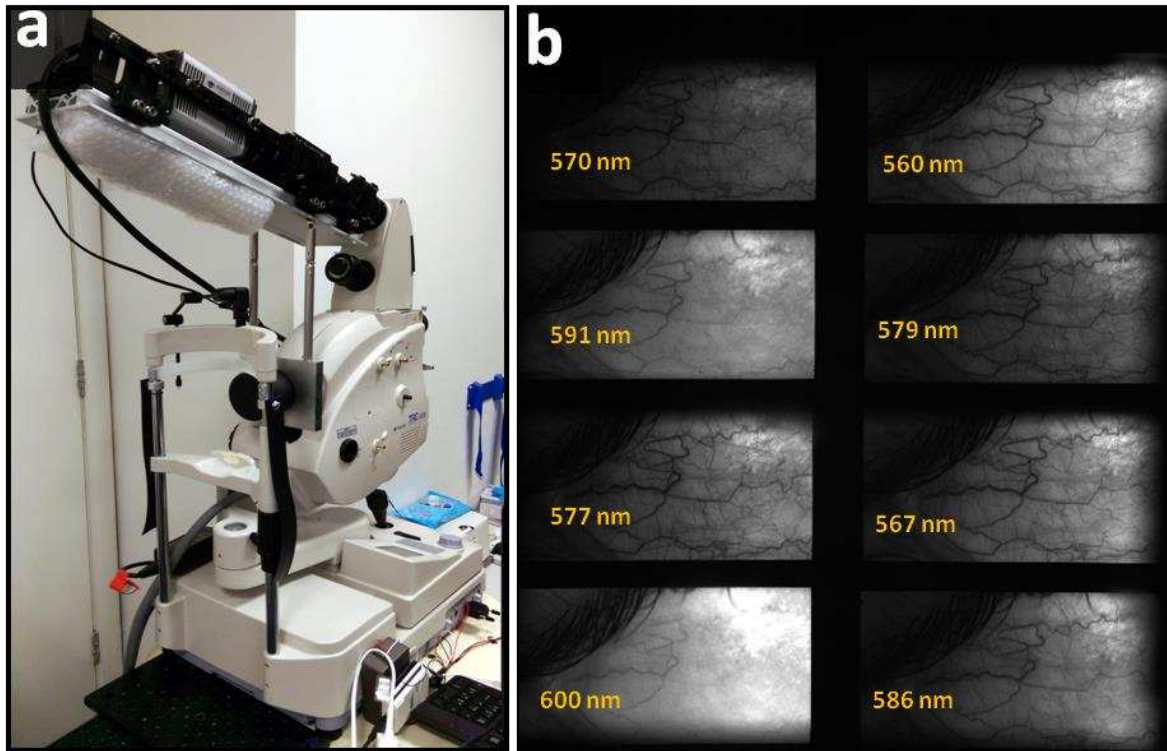
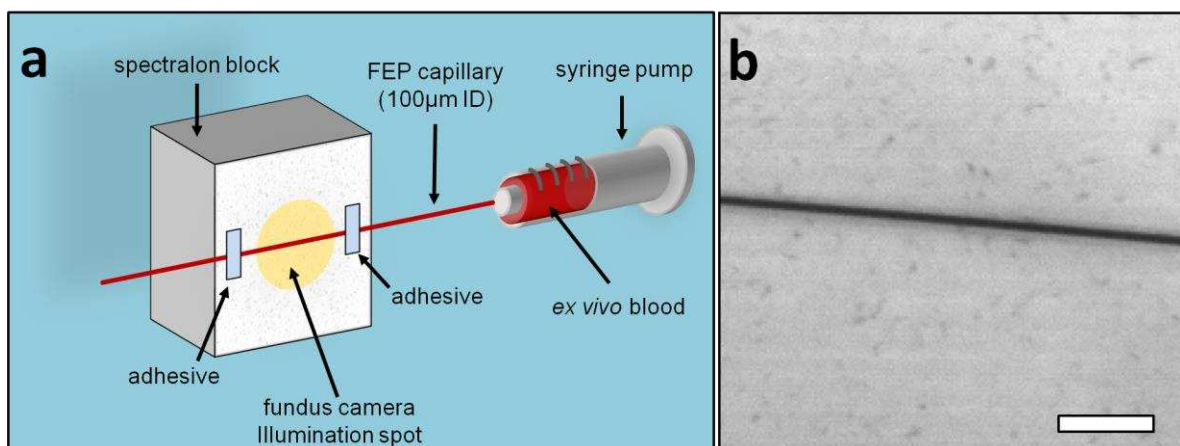
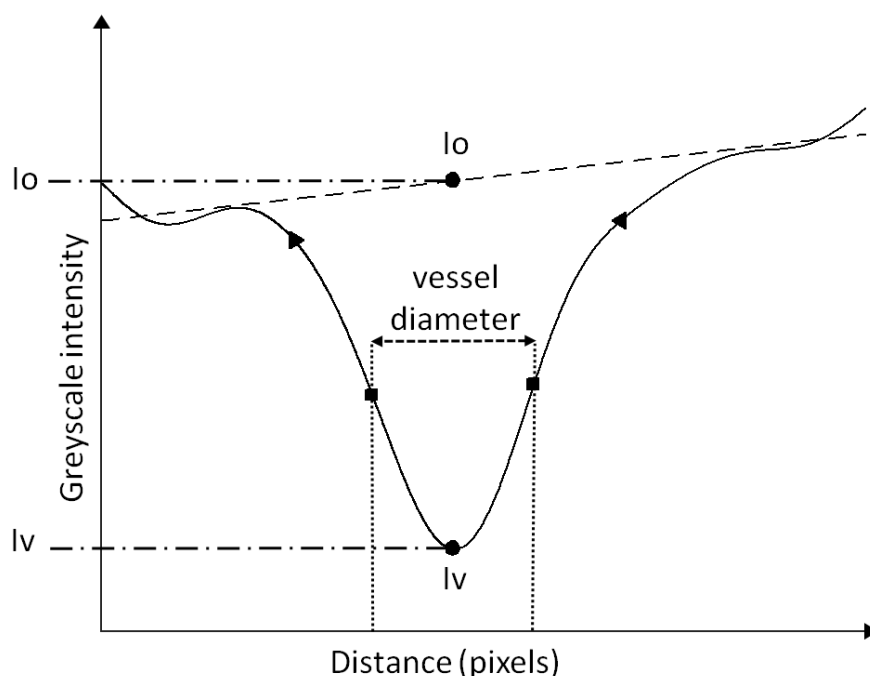


Figure 3. (a) diagram of the scleral phantom. **(b)** 100 μ m capillary filled with blood; scale bar represents one millimetre.



778

779 **Figure 4.** Depiction of the vessel fitting algorithm applied to estimate vessel
 780 diameter, the greyscale intensity in centre of vessel (I_v), and the background
 781 greyscale intensity (I_o). Vessel boundaries are defined as the points of maximum rate
 782 of change of greyscale intensity in the vessel profile.



783

784 **Table 1.** Repeatability of optical-density ratio (ODR) measurements for conjunctival
 785 and episcleral vessels.

Parameter	Bulbar Conjunctival vessels	Episcleral vessels
Number of subjects	10	7
Total number of sampled vessel sections	57	22
ODR repeatability: individual vessels*	2.27%	2.28%
ODR repeatability**	0.96%	1.55%

*standard deviation of 8 repeated measurements of individual vessels, averaged across all subjects

** standard deviation of the average ODR of vessels when averaged by vessel type, then averaged across all subjects

786

Table 2. Average optical-density ratio (ODR), diameter of vessels, and pulse oximeter data at normoxia and hypoxia.

Parameter	Number of subjects	Number of vessel sections analysed	Normoxia	Hypoxia	p-value*
Conjunctival ODR (mean ± SE)	10	64	0.846 ± 0.014	0.916 ± 0.011	<0.001
Episcleral ODR (mean ± SE)	7	24	0.880 ± 0.019	0.938 ± 0.018	0.03
Conjunctival diameter (µm) (mean ± SD)	10	64	80.1 ± 7.6	80.6 ± 7.0	0.89
Episcleral diameter (µm) (mean ± SD)	7	24	78.9 ± 8.7	97.6 ± 14.3	0.02
Fingertip pulse oximeter SO₂ (%) (mean ± SD)	10	N/A	97.1 ± 1.7	86.7 ± 4.3	<0.001

*Pairwise t-test

SE = standard error

SD = standard deviation

Figure 5. Phantom validation; optical-density ratio (ODR) was measured to be inversely proportional to SO_2 as measured by a blood gas analyser (BGA). Vertical error bars represent standard deviation of ODR as measured along the length of the FEP capillary, horizontal error bars represent the blood gas analyser manufacturers quoted error of $\pm 1.8\%$ SO_2 . Dashed line is fitted linear trend ($R^2 = 0.89$).

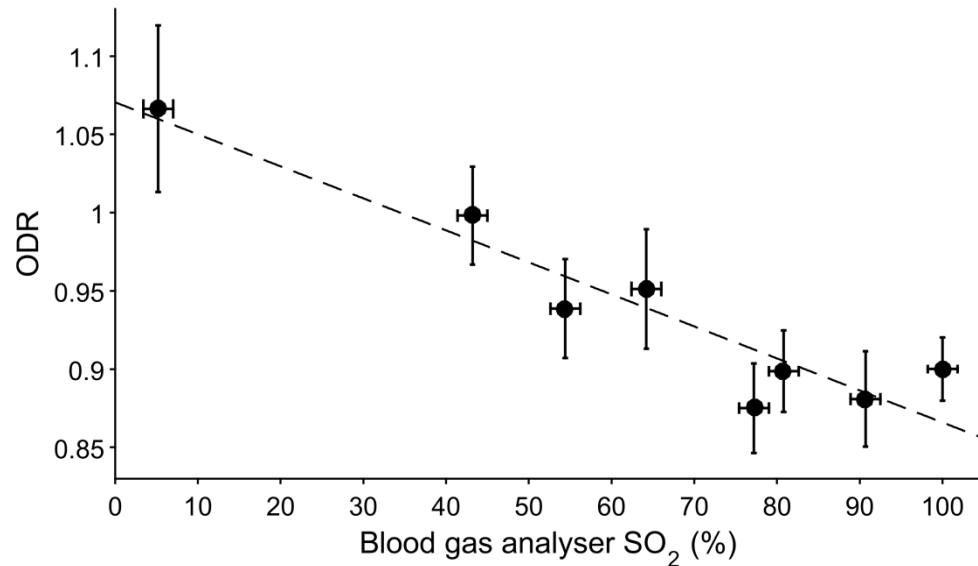
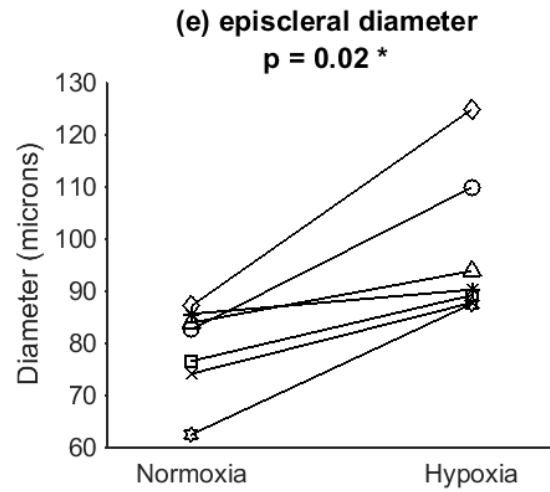
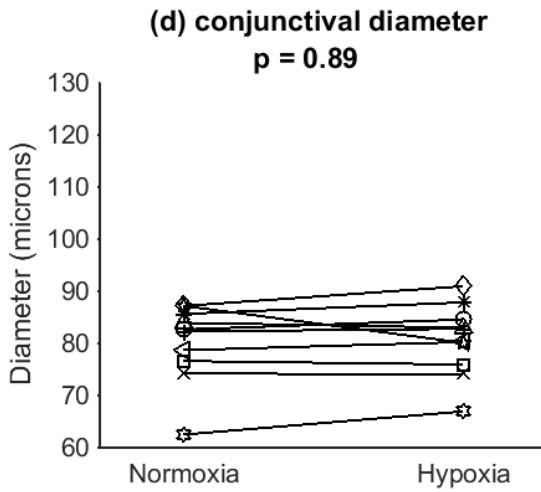
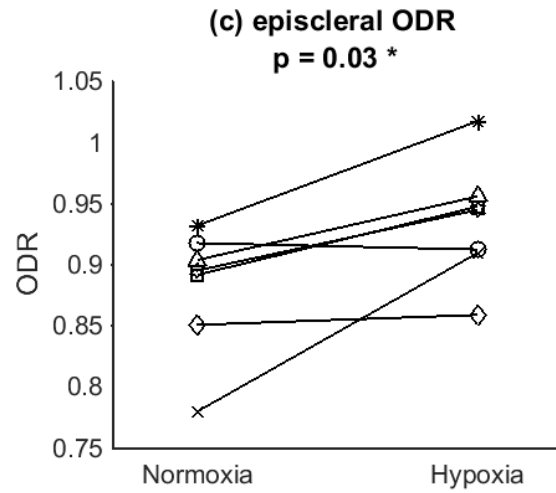
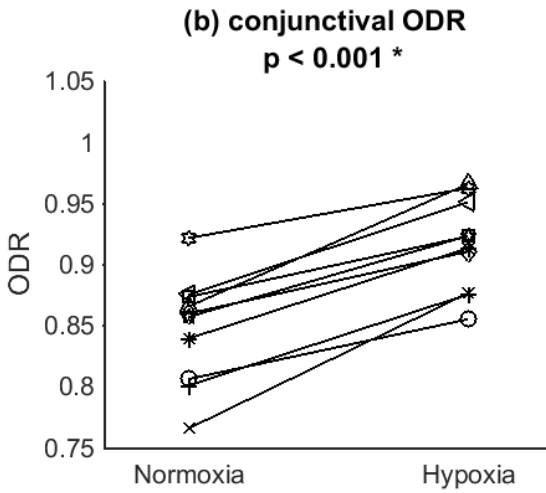
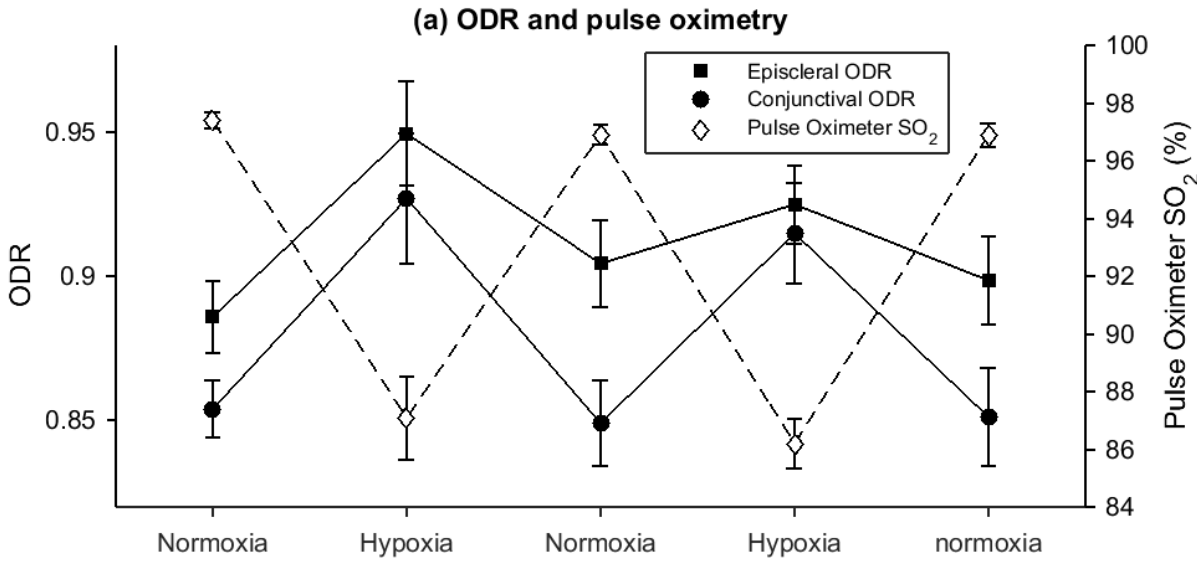
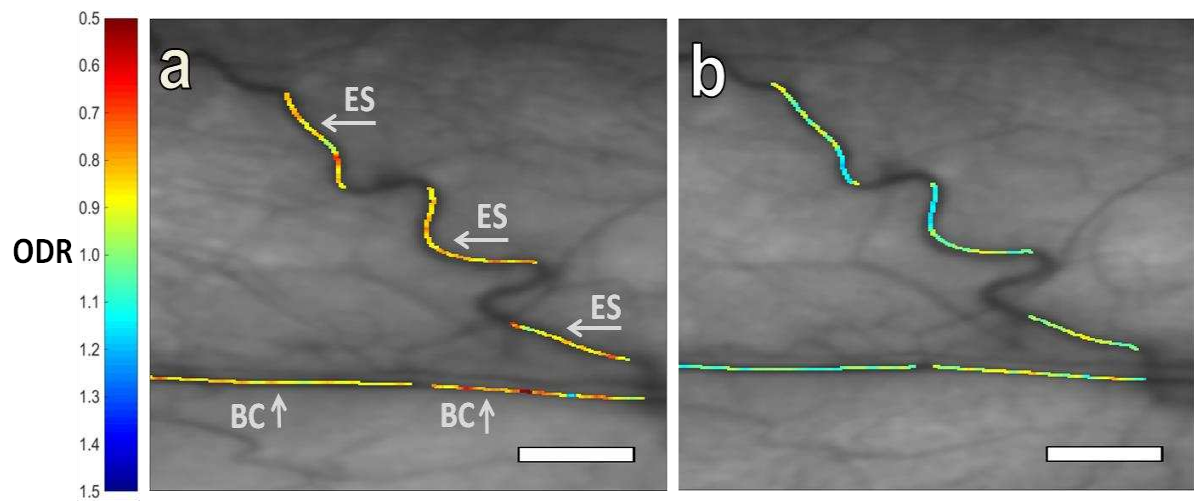


Figure 6. (a) Average optical-density ratio (ODR) and pulse oximeter data throughout the normoxia/hypoxia sequence. Error bars are the standard error of the mean. Graphs **(b)-(e)** show pairwise change of average vessel diameter and average ODR for each subject at normoxia and hypoxia. Statistically significant results are denoted with an asterisk (*).



809 **Figure 7.** Optical-density ratio (ODR) map of vasculature at **(a)** normoxia and **(b)**
810 hypoxia. ODR increases (i.e. SO_2 decreases) with hypoxia. Episcleral vessels are
811 labelled with (ES) and bulbar conjunctival vessels are labelled (BC). Scale bar
812 represents 500 μm .

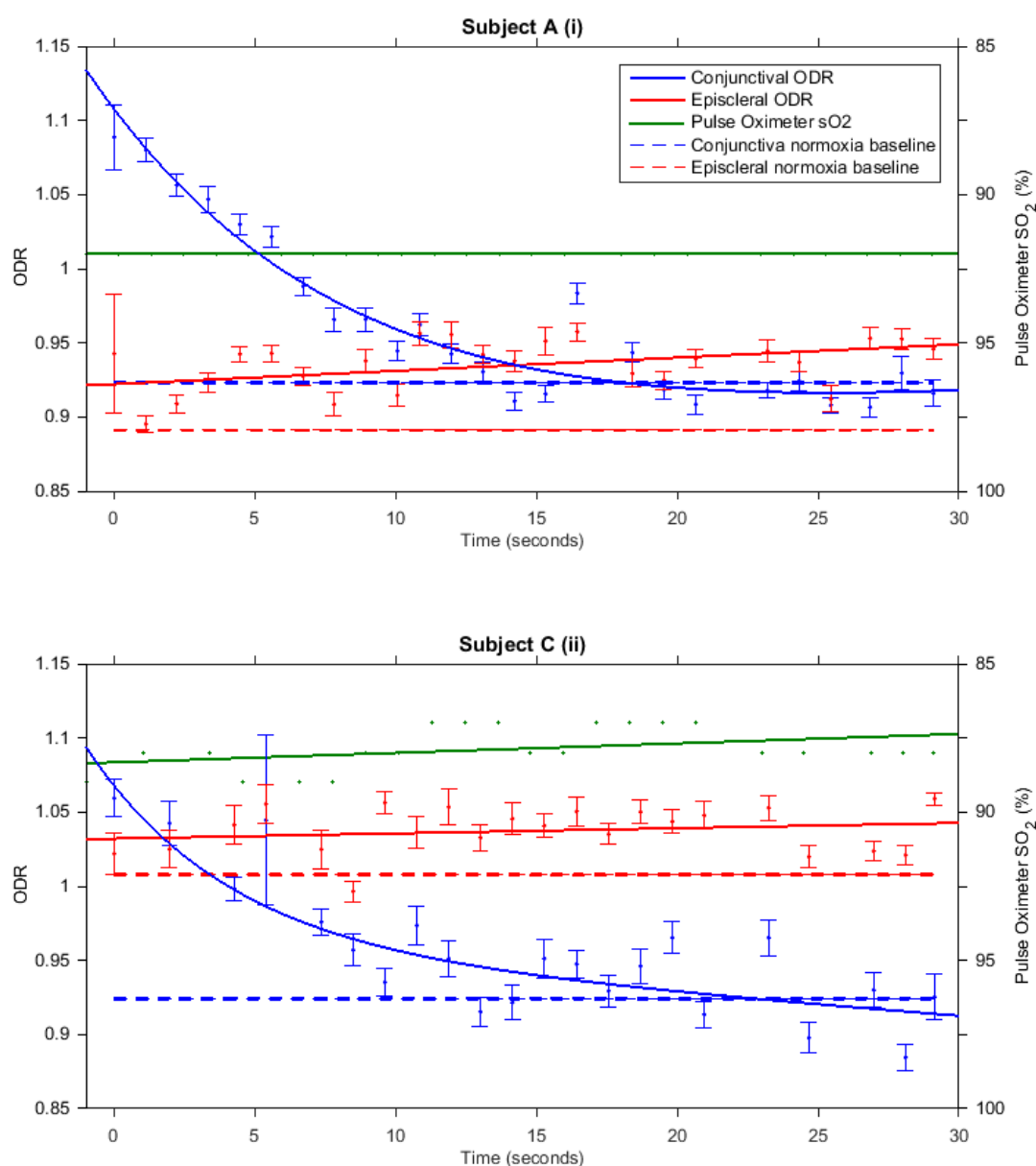


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816 **Figure 8.** Optical-density ratio (ODR) of hypoxic vasculature versus time after eyelid
 817 opening (i.e. exposure to ambient air) in two representative subjects. Bulbar
 818 conjunctival ODR (blue fitted line) decreased exponentially upon eyelid opening
 819 before reaching normoxia baseline levels (blue dashed line). Episcleral ODR (red
 820 fitted line) remained higher than normoxia levels (red dashed line). This indicates
 821 that hypoxic bulbar conjunctival vessels rapidly reoxygenated by oxygen diffusion
 822 when exposed to ambient air whereas hypoxic episcleral vessels (embedded in
 823 episcleral tissue) did not reoxygenate. Error bars represent the standard error of the
 824 mean. The green fitted line is pulse oximeter data ($\pm 2\%$ SO_2 uncertainty quoted by
 825 the manufacturer not depicted for clarity).



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828 **Table 3.** Calculated values of '1/2 time to reoxygenation' ($T_{1/2}$) for 4 subjects,
 829 repeated twice per subject.

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Subject	Data set	$T_{1/2}$ (seconds)
A	(i)	6.6
	(ii)	4.1
B	(i)	3.0
	(ii)	2.9
C	(i)	2.1
	(ii)	3.4
D	(i)	2.2
	(ii)	3.2
Average		3.4
Standard Deviation		1.4

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